

Waters

Transferring Yesterdays Methods
to Tomorrows Technology:
The Evolution from HPLC to UPLC™



Eric S. Grumbach

Thomas E. Wheat

Chuck Phoebe

Joe Arsenault

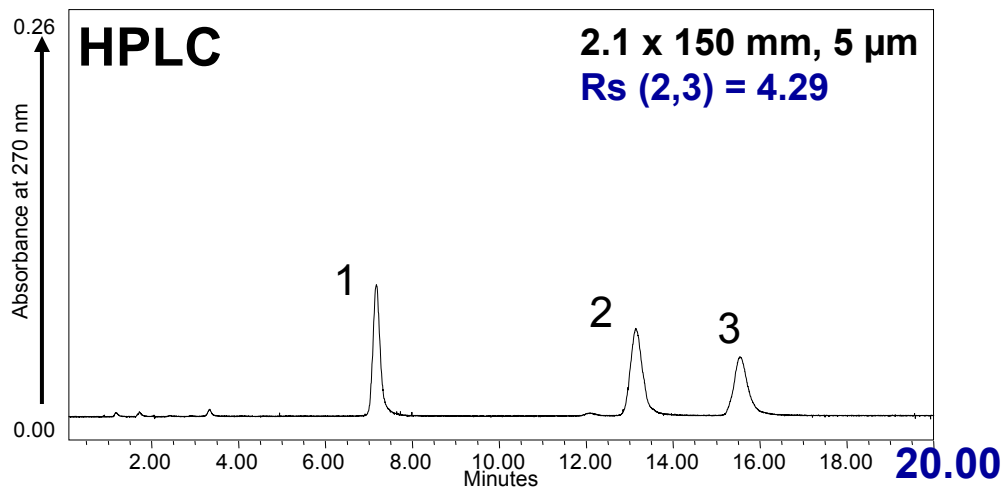
Jeffrey R. Mazzeo

Diane M. Diehl



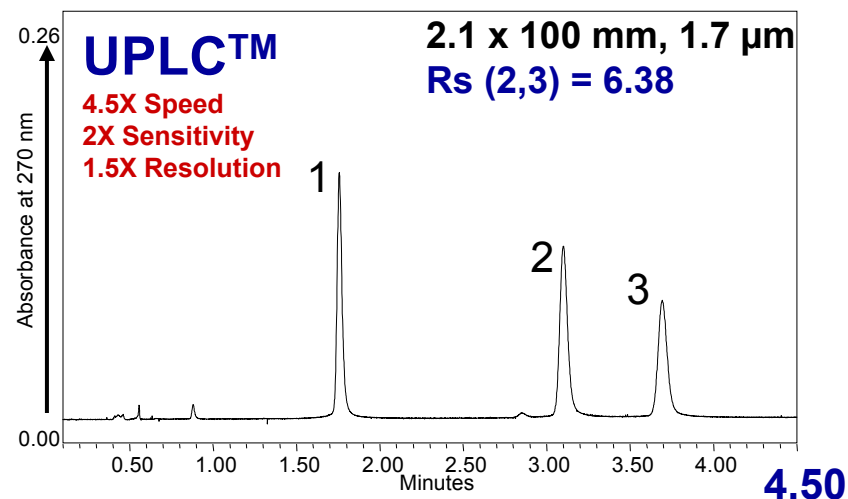
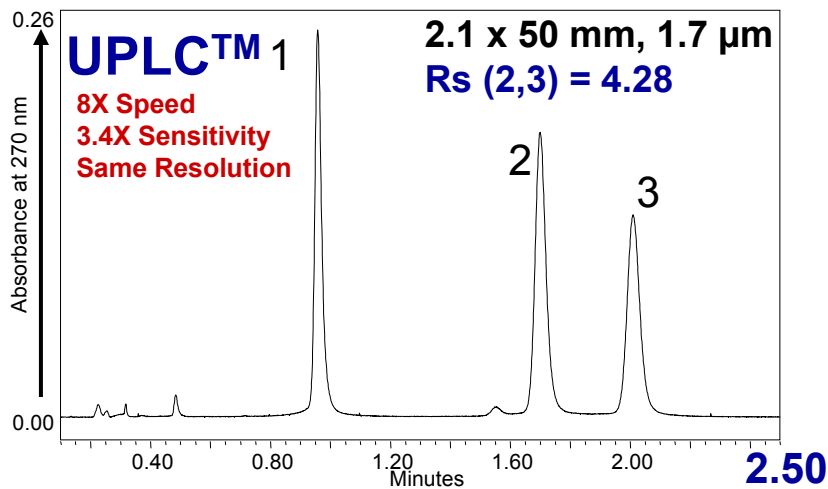
For Complete ☼ Confidence

- A new class of separation science
 - Based on chromatography columns with very small particles
 - Based on instruments designed to take advantage of the small particles
- Provides improved Resolution, Speed, and Sensitivity without compromise
- Suitable for chromatographic applications in general
 - Appropriate for developing new methods
 - Appropriate for improving existing methods



Faster, More Sensitive Methods

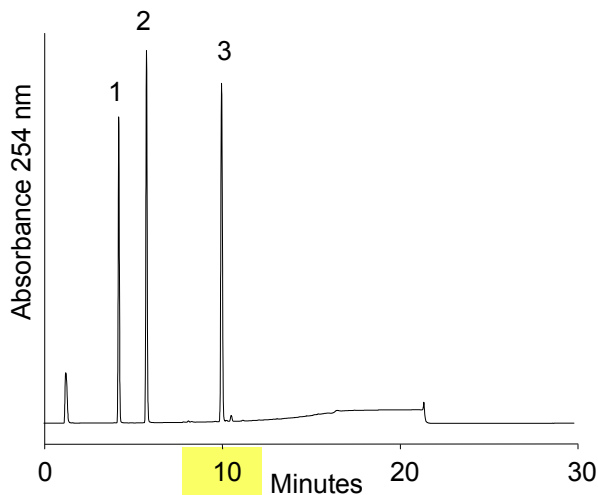
**Faster, More Sensitive,
Higher Resolution Methods**



Choices available:

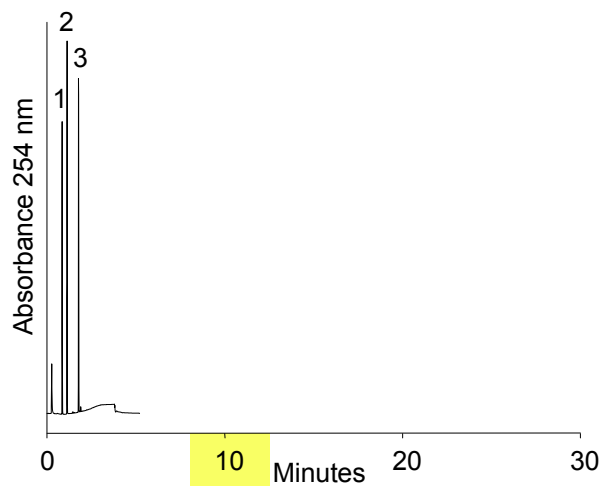
- Increased **speed** and sensitivity with the same resolution
- Increased **sensitivity** and speed with resolution
- Increased **resolution** and sensitivity at the same speed





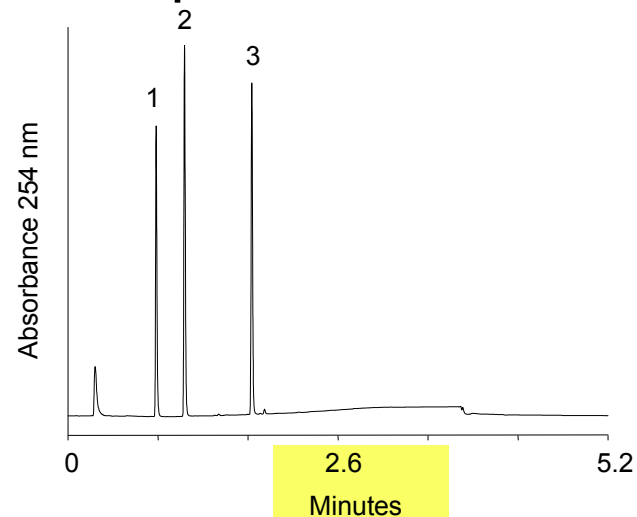
HPLC

An Example of a Successful Method Transfer



Optimized UPLC™

Magnification of Optimized UPLC™



- Classes of methods transfer
 - Replace a column brand with another
 - Replace an instrument with another
 - Replace both the instrument and the column
- The benefits of UPLC™ can only be realized with a new column on a new instrument
- As with any project to transfer a Liquid Chromatographic method, proper consideration of all the factors which control the separation process is essential for success

- **Internal diameter**
 - 2.1 mm or 1.0 mm
- **Length**
 - If primary goal is **speed**, choose 50 mm length
 - If primary goal is **resolution**, choose 100 mm length

<u>L/dp</u>		<u>RATIO</u>		<u>Typical</u> <u>Run Times</u>
$\frac{300\text{mm}}{10\ \mu\text{m}}$	=	30,000	1970's	~ 30+ min.
$\frac{150\text{mm}}{5\ \mu\text{m}}$	=	30,000	1980's	~10-15 min.
$\frac{100\text{mm}}{3\ \mu\text{m}}$	=	33,300	1990's	~ 5-10 min.
$\frac{50\text{mm}}{1.7\ \mu\text{m}}$	=	29,500	2004	~1- 2 min.
$\frac{100\text{mm}}{1.7\ \mu\text{m}}$	=	58,820	2x Maximum Resolution Capability	

If you keep the L/dp ratio the SAME for 2 columns, you will obtain the SAME Resolution. With smaller particle sizes in shorter columns, you will achieve the same separation, but in LESS TIME!

Available Ligands:

- ACQUITY UPLC™ BEH C₁₈
 - Straight chain alkyl C₁₈
- ACQUITY UPLC™ BEH Shield RP₁₈
 - Embedded polar group (carbamate)
- ACQUITY UPLC™ BEH C₈
 - Straight chain alkyl C₈
- ACQUITY UPLC™ BEH Phenyl

Why Multiple Ligands?:

- Changes in hydrophobicity
- Changes in silanol activity
- Changes in hydrolytic stability
- Changes in ligand density
- Changes in selectivity



- Solvent delivery
 - Scale flow rate appropriate to column dimensions
 - Scale linear velocity appropriate to particle size
 - Adjust all segments of method, including equilibration
- Sample injection
 - Scale injection volume
- Detection
 - Ensure proper data sampling rate
 - Ensure proper time constant

4.6 x 150 mm, 5 μ m HPLC Column

Gradient Step	Time Since Injection	Flow Rate	%A	%B	Curve
Initial	0	1.5	95	5	*
2	15	1.5	5	95	6
3	20	1.5	5	95	1
4	30	1.5	95	5	1

- Express gradient duration in % change per column volume (cv) units
- Calculate each segment as a number of column volumes
- Calculate time required to deliver the same number of column volumes to the UPLC™ column at the chosen flow rate.

For 15 min at 1.5 mL/min on a 4.6 x 150 mm column

$$\text{Gradient Volume} = \text{Flow Rate} \times \text{Time} = 1.5 \text{ mL/min} \times 15 \text{ min} = 22.5 \text{ mL}$$

$$\text{Column Volume} = \pi \times r^2 \times L = 3.14 \times 2.3^2 \times 150 = 2.49 \text{ mL}$$

$$\text{Gradient Duration (cv)} = \frac{\text{Gradient Volume}}{\text{Column Volume}}$$

$$\text{Gradient Duration} = \frac{22.5 \text{ mL}}{2.49 \text{ mL}} = 9.03 \text{ cv}$$

4.6 x 150 mm, 5 μ m HPLC Column

Gradient Step	Time Since Injection	Flow Rate	%A	% B	Curve	Segment Duration (min)	Segment Duration (Col.Vol.)
Initial	0	1.5	95	5	*	0	0
2	15	1.5	5	95	6	15	9.03
3	20	1.5	5	95	1	5	3.01
4	30	1.5	95	5	1	10	6.02

- Consider 1.7 μm target particle (2.1 mm i.d. column)
- Assume temperature and viscosity transferred
- Adjust flow rate based on van Deemter curve and approximate molecular weight
 - ~0.6 mL/min for average 500 dalton (molecular weight) molecules
 - ~0.1 mL/min for larger molecules because diffusion is slower, e.g., ~2,000 dalton peptides

Original Step 2: 15 min @ 1.5 mL/min with Duration of **9.03cv**

Calculate Target Step 2: (keeping duration @ **9.03cv**)

$$\text{Target Column Volume } (2.1 \times 50) = 0.17 \text{ mL}$$

$$\text{Gradient Step Volume} = \text{Duration (cv)} \times \text{Target Column Volume}$$

$$= 9.03\text{cv} \times 0.17 \text{ mL} = 1.54 \text{ mL}$$

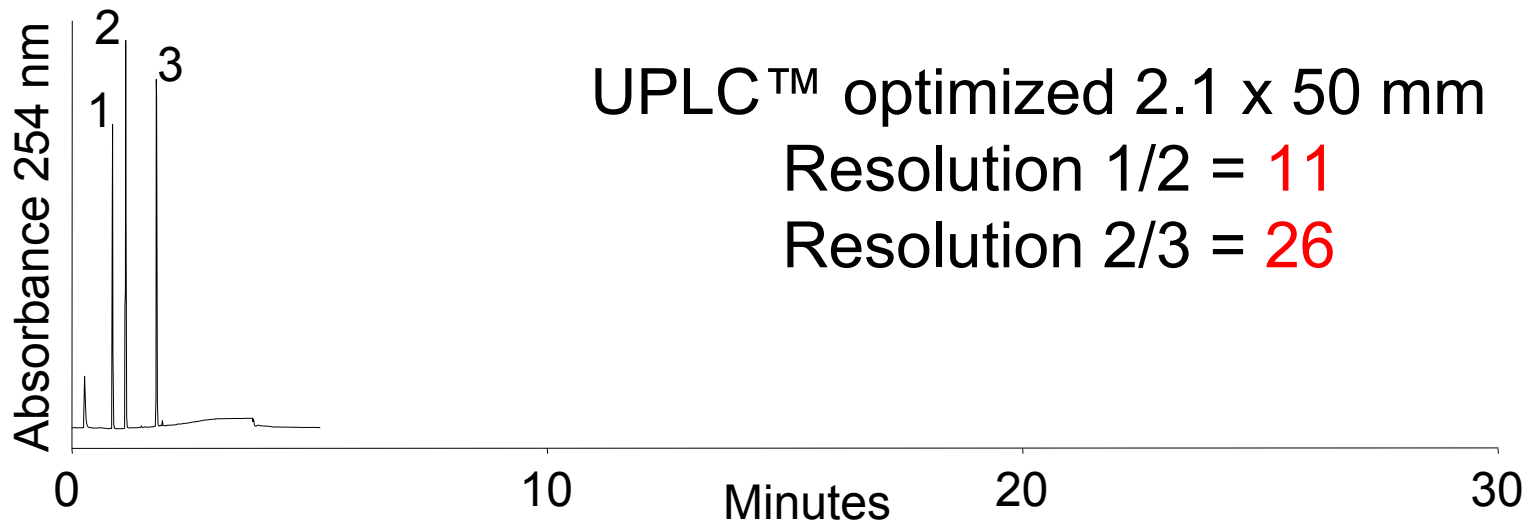
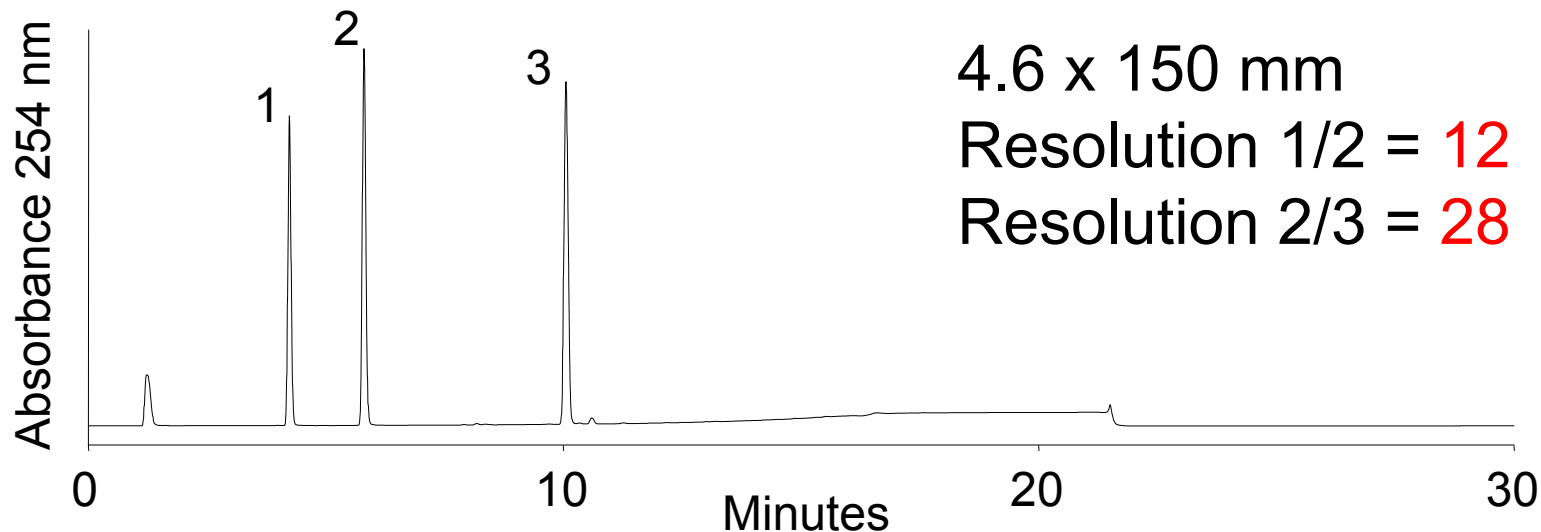
$$\text{Gradient Step Time} = \text{Gradient Step Volume} / \text{UPLC™ Flow Rate}$$

$$= 1.54 \text{ mL} / 0.60 \text{ mL/min.} =$$

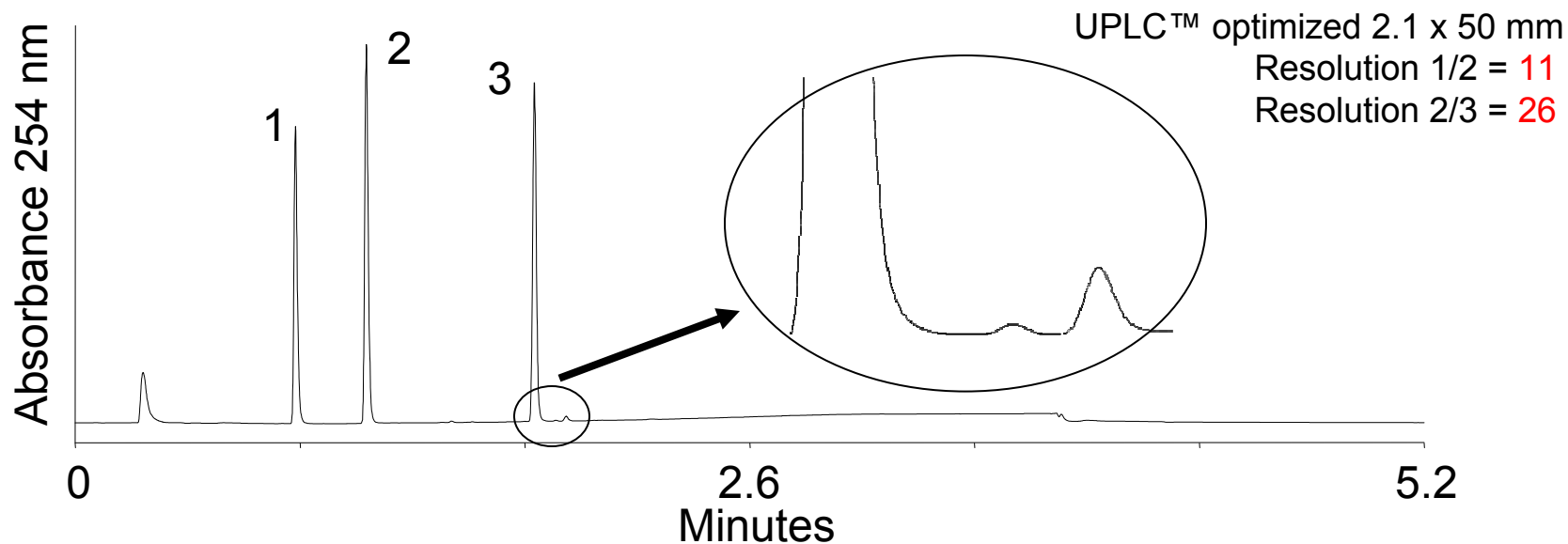
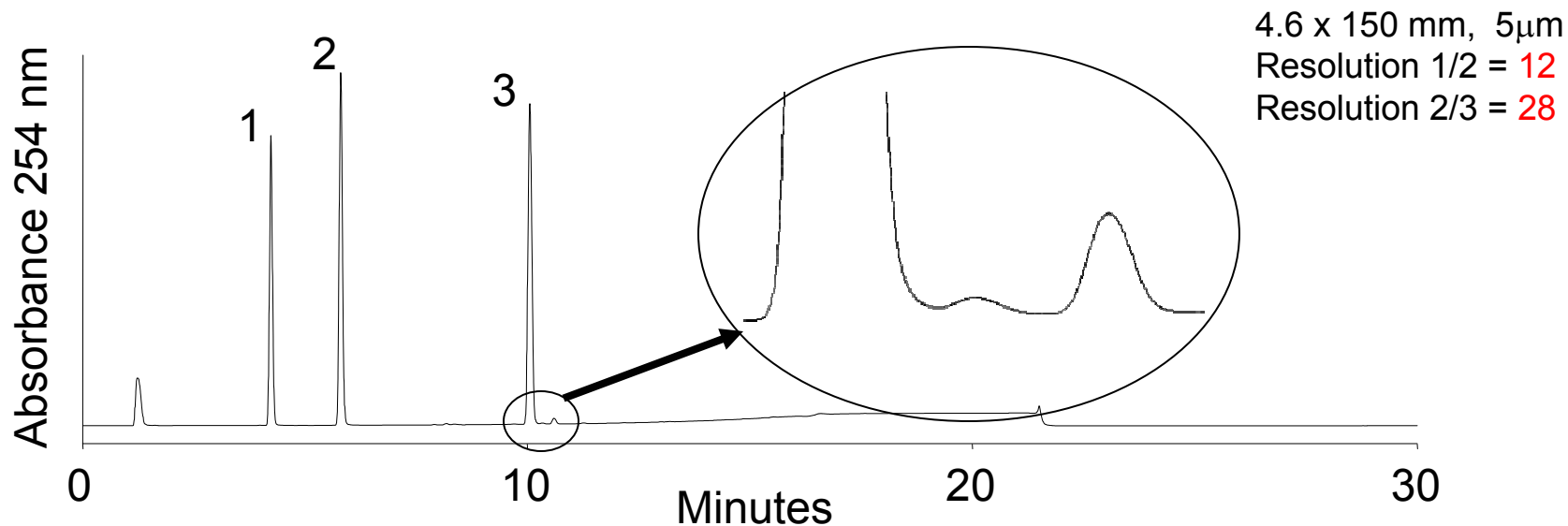
$$\mathbf{2.6 \text{ min.}}$$

2.1 x 50 mm, 1.7 μ m UPLC™ Column

Gradient Step	Time Since Injection	Flow Rate	% A	% B	Curve	Segment Duration (min)	Segment Duration (Col.Vol.)
Initial	0	0.6	95	5	*	0	0
Initial Hold	0	0.6	95	5	1	0	0
2	2.61	0.6	5	95	6	2.61	9.03
3	3.48	0.6	5	95	1	0.87	3.01
4	5.22	0.6	95	5	1	1.74	6.02



Comparing Results: Does the new method meet resolution criteria?



Original HPLC Method: Caffeic Acid Derivatives in Echinacea Purpurea

Chromatographic Conditions :

Columns: XTerra® MS C₁₈ 4.6 x 150 mm, 5.0 µm

Mobile Phase A: 0.1% CF₃COOH in H₂O

Mobile Phase B: 0.08% CF₃COOH in ACN

Flow Rate: 1.0 mL/min

Gradient:	Time	Profile		Curve
	(min)	%A	%B	
	0.0	92	8	6
	2.0	92	8	7
	32.0	50	50	6
	35.0	10	90	6
	36.0	92	8	6
	41.0	92	8	6

Injection Volume: 10.0 µL

Weak Needle Wash: 0.1% CF₃COOH in 8% ACN

Sample: Caffeic Acid Derivatives in Echinacea Purpurea

Sample Diluent: 50:50 H₂O: MeOH with 0.05% CF₃COOH

Sample Concentration: 100 µg/mL

Temperature: 40 °C

Detection: UV @ 330 nm

Sampling rate: 10 pts/sec

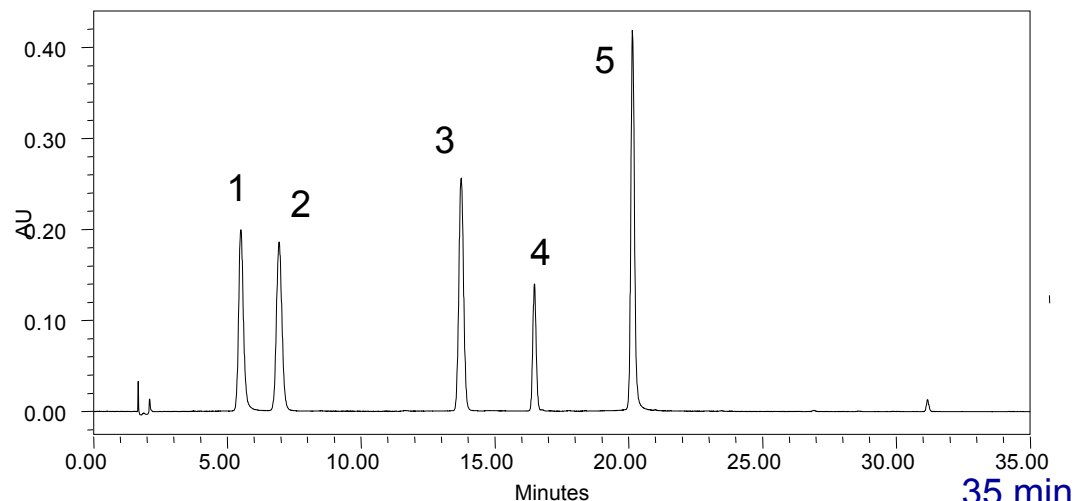
Time Constant: 0.1

Instrument: Alliance® 2695 Separations Module with
2996 PDA detector

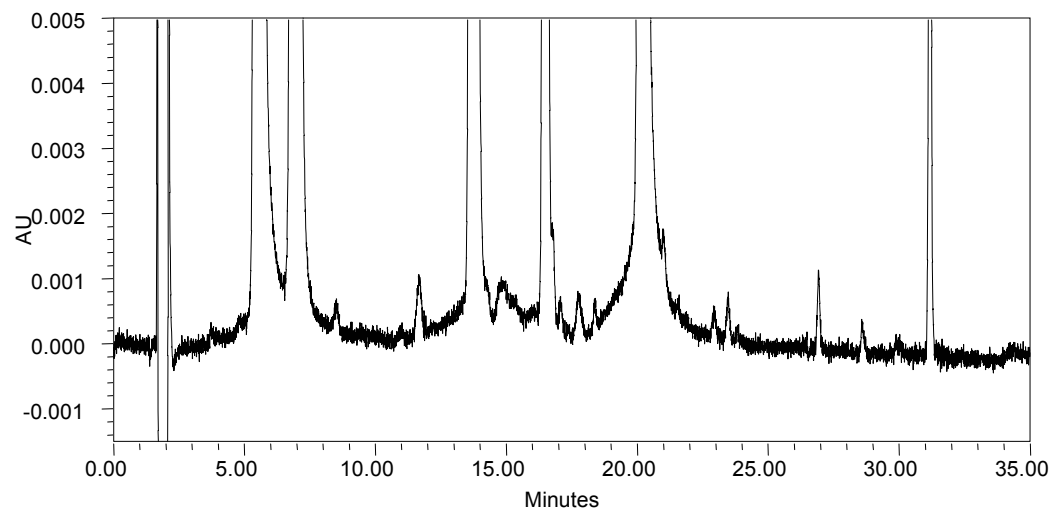
Analyte

1. Caftaric acid
2. Chlorogenic acid
3. Cynarin
4. Echinacoside
5. Cichoric acid

E.G.



35 min.



Enhanced Baseline

Original HPLC Method



Final UPLC™ Method

Chromatographic Conditions :

Columns: XTerra® MS C₁₈ 4.6 x 150 mm, 5.0 µm

Mobile Phase A: 0.1% CF₃COOH in H₂O

Mobile Phase B: 0.08% CF₃COOH in ACN

Flow Rate: 1.0 mL/min

Gradient:	Time	Profile		Curve
	(min)	%A	%B	
	0.0	92	8	6
	2.0	92	8	7
	32.0	50	50	6
	35.0	10	90	6
	36.0	92	8	6
	41.0	92	8	6

Injection Volume: 10.0 µL

Sample: Caffeic Acid Derivatives in Echinacea Purpurea

Sample Diluent: 50:50 H₂O: MeOH with 0.05% CF₃COOH

Sample Concentration: 100 µg/mL

Temperature: 40 °C

Detection: UV @ 330 nm

Sampling rate: 10 pts/sec

Time Constant: 0.1

Instrument: Alliance® 2695 Separations Module with
2996 PDA detector

Chromatographic Conditions :

Columns: ACQUITY UPLC™ BEH C₁₈ 2.1 x 50 mm, 1.7 µm

Mobile Phase A: 0.1% CF₃COOH in H₂O

Mobile Phase B: 0.08% CF₃COOH in ACN

Flow Rate: 0.5 mL/min

Gradient:	Time	Profile		Curve
	(min)	%A	%B	
	0.0	92	8	6
	0.1	92	8	7
	4.45	50	50	6
	4.86	10	90	6
	5.0	92	8	6
	6.0	92	8	6

Injection Volume: 1.0 µL

Weak Needle Wash: 0.1% CF₃COOH in 8% ACN

Sample: Caffeic Acid Derivatives in Echinacea Purpurea

Sample Diluent: 50:50 H₂O: MeOH with 0.05% CF₃COOH

Sample Concentration: 100 µg/mL

Temperature: 40 °C

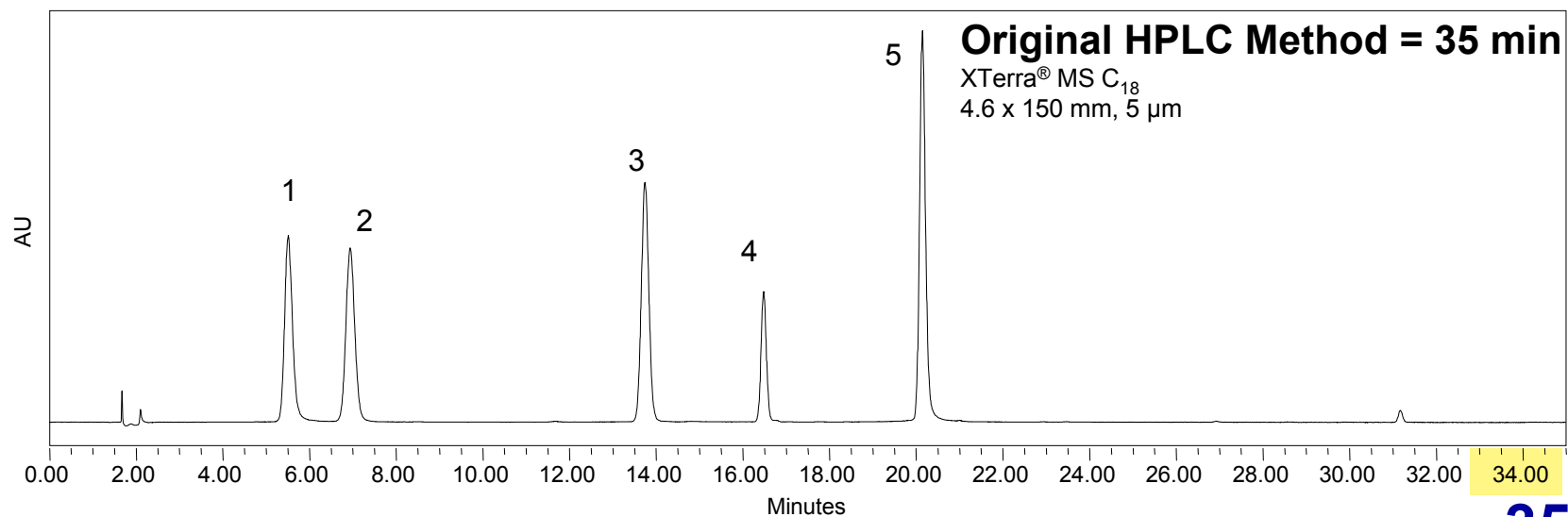
Detection: UV @ 330 nm

Sampling rate: 40 pts/sec

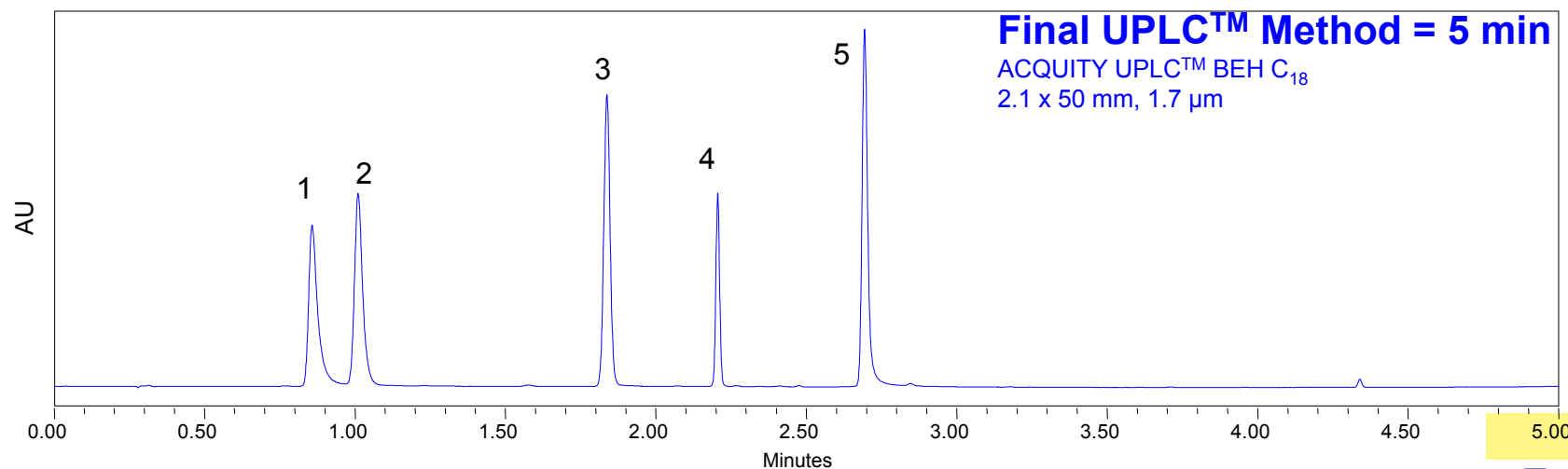
Time Constant: 0.1

Instrument: Waters ACQUITY UPLC™, with TUV detector

HPLC-to-UPLC™ Method Transfer: Caffeic Acid Derivatives in Echinacea Purpurea

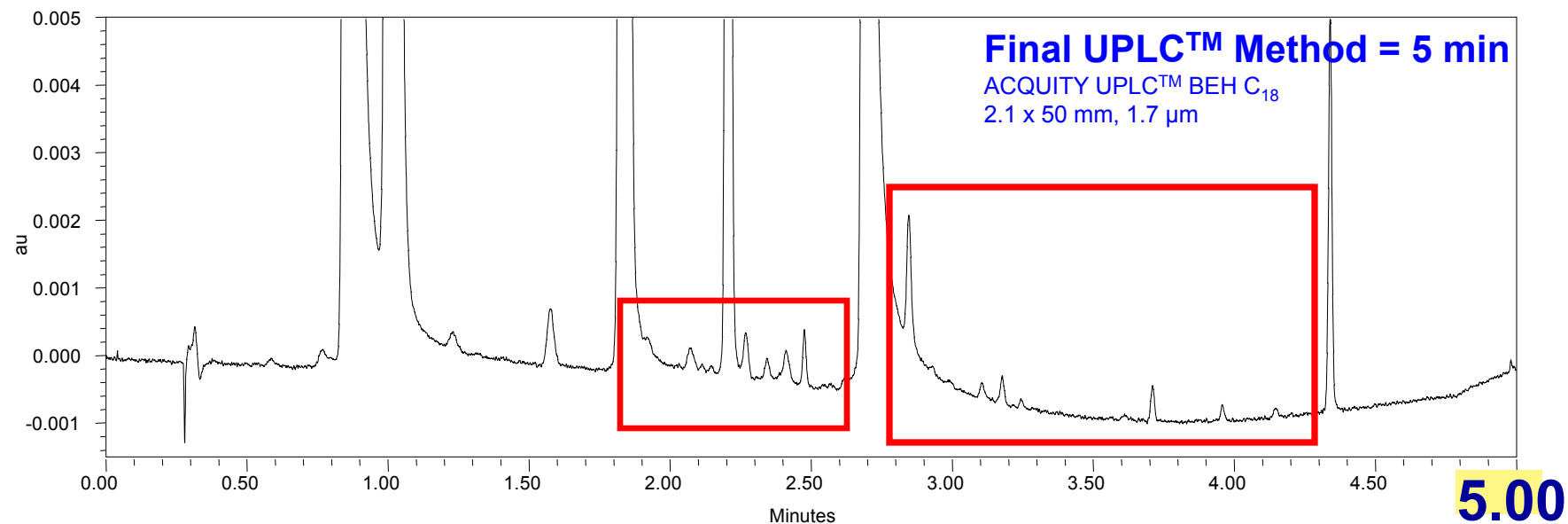
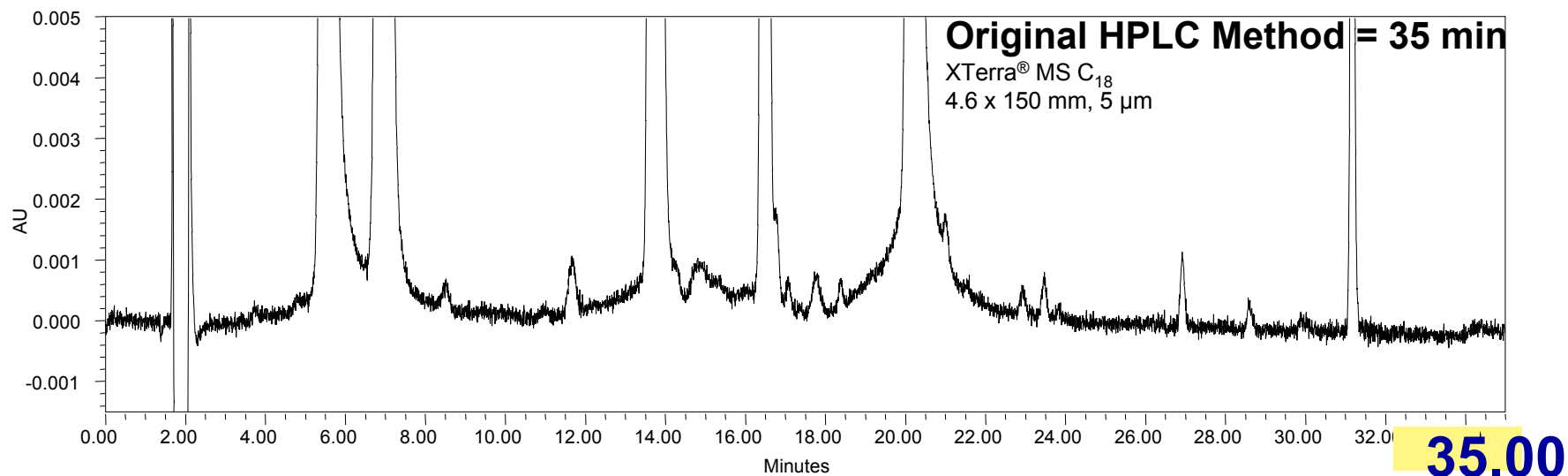


35.00



5.00

HPLC-to-UPLC™ Method Transfer: Caffeic Acid Derivatives in Echinacea Purpurea



- Methods can be moved directly from HPLC to ACQUITY UPLC™
 - Improved resolution
 - Improved speed
 - Improved detectability
- Many parameters can and must be transferred to preserve results
- Attention to detail leads to success