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### CEE 772: Instrumental Methods in Environmental Analysis Lecture #11

### Sample Preparation: Basics and Physical Methods (cont.) (Skoog, nothing)



David Reckhow

CEE 772 #11

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### Pretreatments to LLE

- Acidify to <2</p>
  - 8040: Phenols
- No pH adjustment
  - 8060: Phthalate Esters
  - 8100: PAHs
- Neutralize
  - 8080: Oganochlorine pesticides & PCBs
  - 8090: nitroaromatics & cyclic ketones
  - 8140: organophosphorus pesticides
- Alkaline (>11) then acidify (<2)</li>
  - 8250 & 8270: GC/MS for Semivolatiles
- Drying is sometimes necessary
  - 10 cm column of anhydrous Na<sub>2</sub>SO<sub>4</sub>
  - Addition of powdered anhydrous MgSO<sub>4</sub>



### Soxhlet Extraction



Good for soils, sediments & sludges

Used with thimbles

- Or glass wool plugs

Typical extraction solvents

- Toluene/methanol (10:1)
- Acetone/hexane (1:1)
- Methylene chloride



### **Distillation Theory**



### **Kuderna Danish Concentrators**

- Ideal for concentrating ether extracts
- Common procedures
  - Use with heated water bath
  - Boiling chips
  - Good to pre-wet snyder column by adding 1 mL of solvent to top
  - Balls should "chatter", but chambers should not be flooded
    - May take 10-20 minutes
  - Stop at apparent volume of ~1 mL
  - Exchange solvent may be added at this time & concentration continued



### **Rotary Evaporation**

- Rotary evaporators (also called "rotavaps" in lab slang) are used to remove solvents from reaction mixtures and can accommodate volumes as large as 3 liters.
- A typical rotary evaporator has a heatable water bath to keep the solvent from freezing during the evaporation process. The solvent is removed under vacuum, is trapped by a condenser and is collected for easy reuse or disposal. Most labs use a simple water aspirator vacuum on their rotavaps, so a rotavap can not be used for air and watersensitive materials unless special precautions are taken.





### Use of Rotavaps

- 1. Empty and then replace the solvent collection flask on the unit.
- 2. Place your flask on the rotary evaporator.
- 3. Use the speed control to rotate the flask. A typical rotavap uses a variable speed sparkless induction motor that spins at 0- 220 rpm and provides high constant torque.
- 4. Turn on the aspirator vacuum. On most models, the vacuum on/off control is managed by turning a stopcock at the top of the condenser .
- 5. Lower your flask into the water bath. On most models, a convenient handle (with height locking mechanism) moves the entire condenser/motor/flask assembly up and down. You can also adjust the tilt of the condenser assembly. Be sure not to put the flask into a water bath that exceeds the boiling point of your solvent!! For small amounts of common solvents you don't need to turn on the bath heater.
- 6. The solvent should start collecting on the condenser and drip into the receiving flask. Some solvents (such as ether or methylene chloride) are so volatile that they will also evaporate from the receiving flask and be discharged down the drain. To prevent this you can place a cooling bath on the receiver or (on some models) use a dry ice condenser.
- 7. Once all your solvent has evaporated, release the vacuum, raise the flask out of the water bath and turn off the rotation. Remove your flask.

### Rotavap tips

- Always use distilled water in your heating bath. Otherwise, scale will build up in the bath and coat the thermistor and heating coils. This is very difficult to remove and reduces the efficiency of the bath. In addition, regular tap water will promote the growth of spectacularly disgusting algae colonies, particularly during the summer months.
- To reduce the amount of evaporation from your water bath, simply add some small plastic balls to the water bath. This reduces the surface area for evaporation and therefore the rate at which the water level drops.
- The ground glass joint holding your flask does not need to be greased, but on rare occasions it (or the bump bulb) may get "frozen". Some companies sell special joint clips that can free frozen joints simply by screwing them in one direction.



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## Freeze Dryers in EVE



- Manifold Dryer
  - Labconco Freeze Dryer 8
    - 8 liter
    - Marcus 5c
  - Tray Dryer
    - Labconco
      FreeZone 6
      - 6 liter
      - Marston 24



### **Freeze Drying**



- Three stages
  - Pre-freezing at atmospheric pressure (A)
    - Slow cooling produces large crystals, which improves sublimation
    - Fast cooling better preserves biological samples
  - Primary Drying
    - Pressure is lowered in sample compartment (C) to 0.06 mBar or below
    - Sample pressure drops (B) and sample warms leading to removal of ice by sublimation
    - Rate depends on difference between vapor pressure of the product (B) compared to the vapor pressure of the condenser (D)
      - Condenser is usually 20 C cooler than product (e.g., -50 to -80C)

## Freeze Drying II

- Secondary Drying
  - Removal of last 5-10% of moisture
  - Higher temp: isothermal desorption
  - 30-50% of primary drying time
- Source of heat during drying
  - Ambient (room) heat
    - Manifold drying (Model 8)
    - May be increased with a water bath
  - Thermal conductor shelf
    - Tray drying (FreeZone 6)



DRY CAKE

SAMPLE

INTERFACE

WATER

HEAT

## Tray Dryers

• Typical Sublimation Cycle





### Manifold Dryers

• External valve



### Manifold Dryers

• Flasks



## Manifold Dryers

- Pre-freezing
  - Either shell freeze or angle flask to avoid breakage
    - Water expands when frozen











## Purge and Trap



The purge and trap method is used specifically to remove and concentrate volatile analytes from liquids or solids. The goal in this case is to concentrate 100% of the analyte

Purge gas is swept through the heated sample and volatilized components go with it into a "trap". The trap contains particles made of adsorbent compounds that the analytes adsorb onto.

After trapping the analytes for a specified procedure time, the flow through the trap is reversed and analytes desorb back off of the trap and into the injection port of the GC.

- A. purge & simultaneous trap
- B. thermal desorption





### P&T

- EPA purging vessel design
  - 5 mL sample
  - Room temp.
  - Use  $N_2$  or He
  - 20-40 mL/min
  - 10-15 min



#### PACKING PROCEDURE

CONSTRUCTION

From CFR Title 40



- OV-1
  - Methyl silicone polymer

### Used for EPA methods#

- 8010: Halogenated VOCs
- 8015: Nonhalogenated VOCs (high temp): Ether, Ethanol, MEK
- 8020: Aromatic VOCs (BTEX, chlorobenzenes)
- 8030: (high temp) Acrolein, Acetonitrile, Acrylonitrile

### Purge and Trap

- Commercial Unit
  - Varian LSC-2000



# Closed Loop Stipping

- Brechbuhler unit
  Marston 5
- High concentration factor
  - 1 liter of sample down to 0.010 mL extract



#### PURGE MODE



• Purge and Adsorb on 1.5 mg activated carbon







• Extract into carbon disulfide

### Solid Phase Extraction

- Steps
  - 1. Addition of solvent to solvate the functional groups
  - 2. Adsorption of s step): 1 mL to 1 L
  - 3. Sample clean-u
  - -4. Elution



#### From: Solid Phase Extraction by Thurman & Mills

### SPE cont.

• Three formats: disks, cartridges & syringe barrel



From: Solid Phase Extraction by Thurman & Mills



From: Solid Phase Extraction by Thurman & Mills

### SPE cont.

• Syringe barrel method with vacuum manifold



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 Other methods of sample application to syringe barrels

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Centrifugation

**Positive Pressure** 

### • Disk methods



Figure 11.5. Procedure for using Empore disk, SPEC-47, Novo-Clean disk, or the Speedisk for analysis of water samples using a reversed-phase sorbent.



SP

From: <u>Solid Phase</u> <u>Extraction</u> by Thurman & Mills



### SPE cont.

• Manifold assembly for disk extractions



#### From: Solid Phase Extraction by Thurman & Mills

# SPME



SPME is used to extract from liquid, air, or sludge without using any solvents. A silica fiber coated with a stationary phase for a GC is attached to a syringe. The fiber is exposed to sample for a certain time to allow the phase to become saturated with analyte.

After sampling, the fiber is retracted into the syringe and the syringe gets injected into the inlet of the GC. SPME does not remove all of the analyte because it is an equilibrium reaction. It usually can obtain 30-50% of the molecules.

Because the binding to the stationary phase of the fiber is a partitioning reaction, there is an equilibrium involved. Equilibration time for analytes must be obtained using calibration experiments.

## SPME

### • Schematic of a typical setup



#### From: Solid Phase Extraction by Thurman & Mills

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Common Name of Adsorbent	Type	Area (m²/g)	Pore Size (Å)	Method Refs.	Supplier
Chromosorb 102	styrene-DVB	350	90	357	Johns-Manville
Chromosorb 105	polyaromatic	650	500	68, 353	Johns-Manville
Chromosorb 106	polystyrene	750		68, 353	Johns-Manville
Ostion SP-1	styrene-DVB	350	85	109, 124	Laboratory Instr.
Synachrom	ethylvinylbenzene-DVB	570	45	425	Laboratory Instr.
XAD-1	styrene-DVB	100	200	76, 78	Rohm and Haas
XAD-2	styrene-DVB	350	90	78	Rohm and Haas
XAD-4	styrene-DVB	780	50	112, 386	Rohm and Haas
Porapak Q <sup>a</sup>	ethylvinylbenzene-DVB	735	70	68, 337	Waters Associates
XAD-7	methyl methacrylate	450	80	114, 191	Rohm and Haas
XAD-8	methyl methacrylate	140	250	329	Rohm and Haas
Spheron MD	methacrylate-DVB	320		427	Laboratory Instr.
Spheron SE	methacrylate-styrene	70		417	Laboratory Instr.
Tenax	diphenylphenylene oxide	20	720	345, 346	Applied Science
Polyurethane <sup>b</sup>	ester-open pore	0.6	_	87	varied
Polyurethane <sup>b</sup>	amide ester-foam	0.02	_	7, 83	varied
Polypropylene	propylene		_	415	varied
Polyethylene	ethylene			412, 414	varied
A-24 <sup>c</sup>	anion exchange	28		449	Rohm and Haas
Teflon	tetrafluoroethylene		_	375, 377	varied
Chromosorb T	tetrafluoroethylene	5		378	Johns-Manville
Fluoropak 80	tetrafluoroethylene	3	_	378	Fluorocarbon Company
Unspecified <sup>d</sup>	coated supports	low	—	394	varied
Unspecified <sup>e</sup>	bonded phases	f	_	68, 480	varied

Table I. Characteristics of Polymeric Adsorbents Used for Water Analyses

NOTE: DVB denotes divinylbenzene. <sup>a</sup> Other Porapaks (P to T) have surface areas from 300 to 700 m<sup>2</sup>/g. <sup>b</sup> Generally unspecified; many different types are available. <sup>c</sup> Single example of many different kinds. <sup>d</sup> Many possibilities are available. <sup>e</sup> Most are porous silica with C<sub>4</sub> to C<sub>18</sub> alkyl bonded groups. <sup>f</sup> A wide range is available from about 100 to 600 m<sup>2</sup>/g.

From: Junk, 1987

	Sorbent	Structure	Typical Loading			
		Reversed Phase				
on ts	Octadecyl (C-18) Octyl (C-8) Ethyl (C-2) Cyclohexyl Phenyl Graphitized carbon Copolymers	$-(CH_2)_{17}CH_3$ $-(CH_2)_7CH_3$ $-CH_2-CH_3$ $-CH_2CH_2$ -cyclohexyl $-CH_2CH_2CH_2$ -Phenyl Aromatic carbon throughout Styrene-divinylbenzene	17%C 14%C 4.8%C 12%C 10.6%			
r SPE	Cyano (CN) Amino (NH <sub>2</sub> ) Diol (COHCOH) Silica gel Florisil Alumina	Normal Phase $-(CH_2)_3CN$ $-(CH_2)_3NH_2$ $-(CH_2)_3OCH_2CH(OH)CH_2(OH)$ -SiOH $Mg_2SiO_3$ $Al_2O_3$	10.5%C, 2.4%N 6.4%C, 2.2%N 8.6%C  			
hase	Amino (NH <sub>2</sub> ) Quaternary amine Carboxylic acid Aromatic sulfonic acid	Ion Exchangers $-(CH_2)_3NH_2$ $-(CH_2)_3N^+(CH_3)_3$ $-(CH_2)_2COOH$ $-(CH_2)_3$ -Phenyl-SO <sub>3</sub> H	1.6 meq/g 0.7 meq/g 0.4 meq/g 1.0 meq/g			
lills	Wide pore hydrophobic (Butyl) Wide pore ion exchangers	Size Exclusion —(CH <sub>2</sub> ) <sub>3</sub> CH <sub>3</sub> —COOH	5.9%C 12.2%C			

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 Common sorbents used for SPE

From: <u>Solid Phase</u> <u>Extraction</u> by Thurman & Mills

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Methyl, Ci







• Bonded phases









ODS, C18

From: <u>Solid Phase</u> <u>Extraction</u> by Thurman & Mills







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Cl

### SPE cont.

• Ion exchange mechanism for 2,4-D



#### From: Solid Phase Extraction by Thurman & Mills



From: Solid Phase Extraction by Thurman & Mills

![](_page_38_Figure_0.jpeg)

From: <u>Solid Phase Extraction</u> by Thurman & Mills

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CLL //2 #11

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ESA Metabolite

(ionic)

Alachlor

(non-ionic)

- Breakthrough curve
  - Atrazine on C18

![](_page_39_Figure_2.jpeg)

# Simple Hydrophobicity Test

![](_page_40_Figure_1.jpeg)

## Isolation/Fractionation based on Hydrophobicity

**Back elution with Test Water** NaOH Allows recovery of **TOC#4** fractions and check of direct fractionation XAD-8 Desorbable • hydrophobics = TOC#4 **TOC#5** Desorbable mesophilics = DOC NaOH #5 XAD-4 David Reckhow CEE 772 #11 42 To Waste

### Fractionation based on Hydrophobicity II

![](_page_42_Figure_1.jpeg)

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#### **Comprehensive NOM Fractionation** RIDVES 1984 Water Sample **NAA** Filter Hydrophobic Bases Hydrophobic Resin Weak Hydrophobic Acids Amberlite XAD-8 Hydrophobic Neutrals Cation Exchange Resin Hydrophilic Bases MSC-1 Anion Exchange Resin **Duolite** Humic Acid A-7 Amberlite Fulvic Acid XAD-8 David Reckhow 44 Hydrophilic Neutrals Hydrophilic Acids

### **Molecular Size separations**

- Ultrafiltration
  - series vs parallel
  - membrane calibration
- Size Exclusion Chromatography
  - HPSEC vs LC
- Others
  - FFF

![](_page_44_Figure_8.jpeg)

• <u>To next lecture</u>